Development and Validation of a Stability Indicating HPLC-UV Assay



for Leucovorin Calcium Oral Suspension

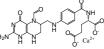
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INTRODUCTION

- Leucovorin, also known as Folinic acid, is used in combination with other chemotherapy agents (mainly methotrexate or 5-fluorouracil) in cancer treatment.
- The objective of this study is to develop and validate a stability indicating HPLC assay for Leucovorin calcium oral suspension.
- The stability of Leucovorin Calcium Oral Suspensions packaged in PTFE plastic multi-dose containers in controlled cold temperature and controlled ambient temperature conditions was evaluated.



METHODS

 Instrumentation: Agilent (Santa Clara, CA) Series 1100 HPLC System equipped with an Agilient Poroshell EC-C8 4.6 x 150 mm 2.7 µm particle column was used for the stability indicating HPLC assay. Barnstead (Lake Balboa, CA) Nanopure Water Systems was used throughout the analysis. HPLC ChemStation Version A.10.02 (Windows XP) software was used for data analysis. Varian (Palo Alto, CA) VMNRS 500 MHz-NMR Spectrometer was used to characterize the Leucovorin standard.

 Materials: 1N Sodium Hydroxide (NaOH) and 1N Hydrochloric Acid from Ricca; Methanol from LabExpress; 30% Hydrogen Peroxide from Sigma-Aldrich; Acetonitrile from Fischer Scientific; Tetrabutylamonium Phydroxide from Spectrum, Leucovorin Calcium standard from USP; 5 mg/mL Leucovorin Calcium Oral Placebo and Leucovorin Calcium Oral Suspension were compounded by Flourish Integrative Pharmacy.

Chromatographic Conditions of stability indicating HPLC method:

Mobile Phase A: 5 mM tetrabutylammonium phosphate buffer solution 80:20 methanol, pH adjusted to 6.6 with tetrabutylammonium hydroxide

Mobile phase B: Methanol.

Gradient: 0-20 min (0-10% B), 20.1-30 min (0% B). Column Temp: 55 $^{\rm o}C;$ Flow Rate: 0.75 mL/min. UV detection at 290 nm.

 Forced degradation studies were performed by subjecting the Leucovorin Calcium oral suspension and placebo to conditions including acid (pH 1 overnight), base (pH 11 overnight), heat (4-hr at 80°C), oxidation (3% H₂O₂) and UV light overnight. Each stressed sample was injected in duplicate. % Leucovorin recovered and retention time of any extra peaks were recorded. Peak purity analysis of the Leucovorin peak in the stressed samples was conducted using a photodiode array detector.

 Robustness: USP standard and samples with Leucovorin and all excipients at 10% of assay level (46.4 µg/mL) were analyzed with the following adjustment to the assay conditions: (+) 2 degree adjustment to the column temperature; (-) 2 degree adjustment to the column temperature; (+) 10% flow rate adjustment; (-) 10% flow rate adjustment; (+) 2% adjustment of methanol in mobile phase; (-) 2% adjustment of methanol in mobile phase; (-) 0.2 unit adjustment of PH in mobile phase;

 The stability of the samples were assessed based on appearance, pH, and potency assay. The samples were stored for 90 days under controlled cold temperature (2°C to 8°C) and controlled ambient temperature (20°C to 25°C) conditions. At time zero a homogeneity study was also conducted where Leucovorin content was determined in each sample bottle by sampling from the top, middle and bottom of the bottles.

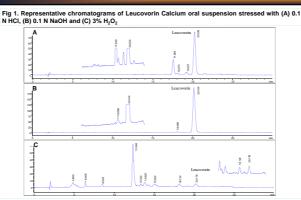


Table 1. Examining the specificity of stability indicating HPLC assay through forced degradation

Sample type	Average Area of Leucovorin Peak (mAU's)	% Of Control	Additional Peaks (RRT)	
Leucovorin Formulation Control	2028.78	N/A	0.87	
Leucovorin Formulation + HCI	705.73	35%	0.11, 0.14, 0.24, 0.63, 0.67, 0.68, 0.73, 0.74, 0.87, 0.89, 0.95, 1.06	
Leucovorin Formulation + NaOH	2007.1	99%	0.26, 0.29, 0.83, 0.87, 0.95	
Leucovorin Formulation + H2O2	89.89	4%	0.24, 0.25, 0.27, 0.32, 0.42, 0.61, 0.66, 0.69, 0.71, 0.74, 0.80, 0.84, 0.86, 0.95	
Leucovorin Formulation + UV	2032	100%	0.26, 0.29, 0.95	
Leucovorin Formulation + Heat/Humidity	2316.32	114%	0.26, 0.29, 0.87, 0.95	

In all conditions, additional peaks were resolved from the API peak. Peak purity of Leucovorin peak was assessed to verify no co-eluting impurities.

Table 2. The accuracy and precision of HPLC assay for Leucovorin Calcium oral suspension

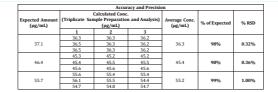
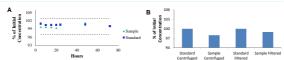


Figure 2. (A) Solution stability and (B) Filter/Centrifuge suitability of Leucovarin standard and assay samples



RESULTS

Table 3. Robustness of Leucovorin Calcium method

Condition Variations	% RSD of Five Standard injections	Tailing Factor	Column Efficiency	Resolution
		(First Std/Last Std/Spl)	(First Std/Last Std/Spl)	(First Std/Last Std/Spl)
Flowrate + 10%	0.1	0.972/0.979/0.979	26555/31020/30218	NA/NA/5.3
Flowrate -10%	1	1.001/1.004/1.012	3910/4057/4054	1.6/1.5/1.6
Column temp + 2*C	0.2	1.001/1.006/1.011	36957/37261/37085	NA/NA/8.0
Column temp - 2 *C	0.1	1.170/1.141/1.130	32788/33001/33483	NA/NA/6.7
pH + 0.2	0.2	0.884/0.897/0.901	20605/21170/22072	NA/NA/3.9
pH - 0.2	0.2	0.928/0.930/0.941	32084/34124/34002	NA/NA/9.9
Organic in MP + 2%	0.2	0.878/0.874/0.878	17129/18124/18997	NA/NA/6.0
Organic in MP - 2%	0.4	0.972/0.984/0.997	2485/2684/2667	1.9/2.1/2.1

Figure 3. (A) Linearity of HPLC assay and (B) homogeneity of Leucovorin Calcium oral suspension in sample bottles

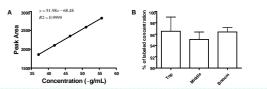


Figure 4. Plot of average Leucovorin recovery relative to the label potency (% of Label) for samples stored at refrigerated and ambient conditions for 90 days. Dashed lines represent lower and upper limits for Leucovorin potency (90-110%).

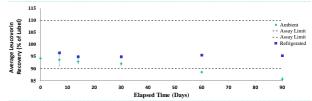
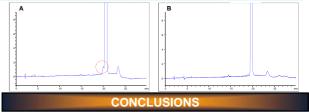


Figure 5. Representative Leucovorin chromatograms with (A) and without (B) artificial peak caused by tetrabutylammonium phosphate during thorough investigation.



 The developed stability-indicating HPLC method enables a quantitative evaluation of the stability of Leucovorin Calcium oral suspension. The validation demonstrates that the method is specific, linear, precise, accurate, and robust.



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